#### VISUALISING THE NANOWORLD:

#### OPTICAL MICROSCOPY AND SUPER RESOLUTION MICROSCOPY

#### Fluorescence

Emission of light by a substance as the results of the absorption of light of lower wavelength.



Example of a fluorescent dye excitation and emission spectra

#### Basic experimental setup - the optical microscope



-> Observe biomolecules, cell components...

**Excitation filter** to cut out any unwanted excitation light, essentially to cut out light from the excitation source that overlaps with the transmission window of the emission filter.

# **Emission filter** to cut out the excitation light.



## Fluorescent markers (dyes)

- GREEN FLUORESCENT PROTEIN
- ORGANIC FLUOROPHORES
- QUANTUM DOTS

## Fluorescent markers

#### **GREEN FLUORESCENT PROTEIN**

- 1960s: isolated from jellyfish Aequorea victoria.
- Since then: many engineered mutants have been developed with better spectral characteristics compared to the wild type GFP (increased fluorescence, better photostability).
- Nobel Prize in Chemistry 2008
- They are produced biologically. Their sequence is known and their DNA can be combined with other proteins to form fusion proteins.





Wavelength (nm)



There are also other variants with other colors: CFP, YFP, RFP

DsRed: protein from *Discosoma* (coral)

http://microscopy.duke.edu

#### Fluorescent markers

#### ORGANIC FLUOROPHORES

- Ex: Fluorescein, Rhodamine, Cy3, Cy5, TRITC, FITC etc...)

![](_page_5_Figure_3.jpeg)

- They can be attached to proteins (including antibodies)

## Fluorescent markers

QUANTUM DOTS: semiconductor nanocrystals-excitons confined in 3 dimensions.

![](_page_6_Figure_2.jpeg)

![](_page_6_Picture_3.jpeg)

Usually made in toxic materials but coatings are preventing any material leakage/ dissolution in bio media (in addition to improving the optical properties of the QD such as yield and stability.)

Suitable for long term studies

# How to label specifically certain parts (proteins, biomolecules) of the cell fluorescently???

![](_page_7_Picture_1.jpeg)

#### Immunofluorescence

Using the molecular recognition between an antibody and an antigen to label fluorescently specific biomolecules

![](_page_8_Figure_2.jpeg)

http://www.apsubiology.org

#### Antigen=Antibody Generator

*Epitope* = part of the antigen that is recognized

by the immune system

Antibodies are proteins synthesized by immune cells: they are used to identify (label) foreign objects in the body.

They can be made against nearly all molecules: Animals produce antibodies in response to antigens recognized as foreign by their immune system.

Antibodies can be labelled with fluorophores and be used to visualize subcellular structures.

## Direct immunofluorescence

![](_page_9_Figure_1.jpeg)

Advantages: fast method, less non specific binding

**Drawbacks:** low sensitivity- limited by the number of fluorophore that can be bound to the primary antibody

![](_page_10_Figure_0.jpeg)

Fixate cells using paraformaldehyde (preserve the tissues by crosslinking proteins) Permeabilize the cells (detergeant)

Prevent non-specific binding (main source = hydrophobic interactions):

-> Add bovine serum albumine or serum (not from the AB target species!) before and during incubation with antibodies.

![](_page_11_Picture_3.jpeg)

Retinal cells stained using DAPI (blue), GFAP (rabbit polyclonal green) and DyLight-488 donkey anti-rabbit  $\beta$ -tubulin III (mouse monoclonal, red) and DyLight-549 donkey anti-mouse

## Confocal microscopy

![](_page_12_Figure_1.jpeg)

#### Confocal microscopy

![](_page_13_Figure_1.jpeg)

### Confocal microscopy

![](_page_14_Figure_1.jpeg)

http://www.uni-mainz.de/FB/Chemie/AK-Janshoff/Illustrationen/ scheme\_confocal\_microscopy.jpg

## 3D reconstruction from confocal images

Nanowires in the rat brain (green: ED1, red: astrocytes, white: nanowires, blue: cell nuclei)

![](_page_15_Picture_2.jpeg)

Linsmeier et al. Nano Lett. 2009

## Optical microscopy: Abbe resolution limit

Due to diffraction a point source of light appears as a finite object described by the point-spread function (PSF).

$$d \sim \frac{\lambda}{2n\sin\alpha} = \frac{\lambda}{2NA} \sim 250nm$$

NA: numerical aperture of the objective n=c/v: refractive index of the medium  $\alpha$ : half-angle of the maximum cone of light that can enter the lens.

![](_page_16_Figure_4.jpeg)

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![](_page_17_Figure_4.jpeg)

![](_page_17_Picture_5.jpeg)

It can be difficult to resolve 2 objects that are close to one another

## Super resolution light microscopy

STORM "Stochastic Optical Reconstruction Microscopy"

= Randomly switch on and off individual fluorophores and localize their position.

(Xiaowei Zhuang- Harvard University)

# **STORM**

![](_page_19_Figure_1.jpeg)

# STORM

![](_page_20_Figure_1.jpeg)

![](_page_20_Figure_2.jpeg)

Xiaowei Zhuang's group-Harvard University

# STORM

![](_page_21_Picture_1.jpeg)

![](_page_22_Figure_0.jpeg)

microtubules

Bo Huang et al, Science 2008

#### STED (Stimulated emission depletion) microscopy

![](_page_23_Figure_1.jpeg)

Confocal microscope with two parallel laser beams. The excitation beam excites the fluorophores and the other beam forces the emission to take place in a color that is not detected through stimulated emission. The result is a smaller point spread function and thereby an improved resolution.

(Stefan Hell, Max Planck Institute, Göttingen) Nobel Prize in Chemistry 2014

#### **STED - Stimulated Emission Depletion**

![](_page_24_Figure_1.jpeg)

**STED - Stimulated Emission Depletion** 

![](_page_25_Figure_1.jpeg)

![](_page_25_Figure_2.jpeg)

#### STED – Stimulated Emission Depletion

![](_page_26_Picture_1.jpeg)

Willig et al Science 2006 Schmidt Nat Methods 2008

#### STED – Stimulated Emission Depletion

![](_page_27_Picture_1.jpeg)

Willig et al Science 2006 Schmidt Nat Methods 2008